

TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371

PU-9843

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR

To be assigned
09/869023

INTERNATIONAL APPLICATION NO.

PCT/EP99/10123

INTERNATIONAL FILING DATE

20 December 1999

PRIORITY DATE CLAIMED

22 December 1998

TITLE OF INVENTION

Removal/Purification of Serum Albumins

APPLICANT(S) FOR DO/EO/US

Tor Regberg and Christel Strom

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (24) indicated below.
4. ☒ The US has been elected by the expiration of 19 months from the priority date (Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371 (c) (2))
 - a. ☒ is attached hereto (required only if not communicated by the International Bureau).
 - b. ☒ has been communicated by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
 - a. ☐ is attached hereto.
 - b. ☐ has been previously submitted under 35 U.S.C. 154(d)(4).
7. ☐ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))
 - a. ☐ are attached hereto (required only if not communicated by the International Bureau).
 - b. ☐ have been communicated by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☐ have not been made and will not be made.
8. ☐ An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).
10. ☐ An English language translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).
11. ☒ A copy of the International Preliminary Examination Report (PCT/IPEA/409).
12. ☒ A copy of the International Search Report (PCT/ISA/210).

Items 13 to 20 below concern document(s) or information included:

13. ☒ An Information Disclosure Statement under 37 CFR 1.97 and 1.98
14. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☒ A **FIRST** preliminary amendment.
16. ☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
17. ☐ A substitute specification.
18. ☐ A change of power of attorney and/or address letter.
19. ☐ A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825.
20. ☐ A second copy of the published international application under 35 U.S.C. 154(d)(4).
21. ☐ A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).
22. ☒ Certificate of Mailing by Express Mail
23. ☒ Other items or information:

duplicate copy of this transmittal letter for charging purposes
copy of the International Application as published by the International Bureau
return postcard

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR 1.53)
To be assigned 09/869023INTERNATIONAL APPLICATION NO.
PCT/EP99/10123ATTORNEY'S DOCKET NUMBER
PU-9843

24. The following fees are submitted:

BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)) :

- ☐ Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO \$1000.00
- ☒ International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$860.00
- ☐ International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$710.00
- ☐ International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$690.00
- ☐ International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) \$100.00

ENTER APPROPRIATE BASIC FEE AMOUNT =**\$860.00**Surcharge of \$130.00 for furnishing the oath or declaration later than ☐ 20 ☐ 30 months from the earliest claimed priority date (37 CFR 1.492 (e)).**\$0.00**

CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE
Total claims	10 - 20 =	0	x \$18.00
Independent claims	2 - 3 =	0	x \$80.00

\$0.00**\$0.00**Multiple Dependent Claims (check if applicable). ☐**\$0.00****TOTAL OF ABOVE CALCULATIONS =****\$860.00**☐ Applicant claims small entity status. (See 37 CFR 1.27). The fees indicated above are reduced by 1/2.**\$0.00****SUBTOTAL =****\$860.00**Processing fee of \$130.00 for furnishing the English translation later than ☐ 20 ☐ 30 months from the earliest claimed priority date (37 CFR 1.492 (f)).**\$0.00****TOTAL NATIONAL FEE =****\$860.00**Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable). ☐**\$0.00****TOTAL FEES ENCLOSED =****\$860.00****Amount to be:****\$****charged****\$**

- a. ☐ A check in the amount of _____ to cover the above fees is enclosed.
- b. ☒ Please charge my Deposit Account No. 500-588 in the amount of \$860.00 to cover the above fees. A duplicate copy of this sheet is enclosed.
- c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 500-588. A duplicate copy of this sheet is enclosed.
- d. ☐ Fees are to be charged to a credit card. **WARNING:** Information on this form may become public. **Credit card information should not be included on this form.** Provide credit card information and authorization on PTO-2038.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

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SIGNATURE

Royal N. Ronning, Jr.

NAME

32,529

REGISTRATION NUMBER

June 20, 2001

DATE

PU-9843

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Application of: T. Regberg, et al. Group Art Unit: To be assigned
Serial Number: To be assigned Examiner: To be assigned
Filing Date: June 20, 2001
Title: Removal/Purification of Serum Albumins

FIRST PRELIMINARY AMENDMENT

Honorable Assistant Commissioner of Patents
Box New Patent Application
Washington, D.C. 20231

Sir:

Please consider the following amendments and remarks in connection with the prosecution of the captioned application, which is a filing under 35 U.S.C. § 371 and claims priority to international application number PCT/EP99/10123 filed December 20, 1999 and Swedish application number 9804465.4 filed December 22, 1998.

In the Claims

Please amend page 18, line 1 as follows:

[C L A I M S]

What is claimed is:

Please amend claim 1 as follows:

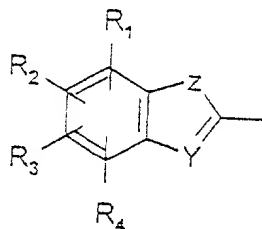
1. (once amended) [A]In a method for selectively enriching/removing a serum albumin from a mixture of other compounds by contacting said mixture with a ligand (= X), the improvement comprising said ligand
 - a) having affinity for and enabling binding of the serum albumin and

- b) being attached via a spacer (= B) to a base matrix (= M') insoluble in the aqueous media used, the matrix with the attached ligand being represented by



where M is the matrix, B is the spacer and X the affinity ligand, with the provision that M may contain further groups X linked via a spacer,

characterized in that said ligand X has been selected among serum albumin-binding structures complying with the formulae



in which

- the free valence bind to the spacer B;
- R₁₋₄ are selected from hydrogen, electron-withdrawing groups, such as halogens and lower alkyl groups (C₁₋₁₀) that possibly are substituted with electron withdrawing groups, such as halogens;
- Z and Y are selected among oxygen, sulphur or nitrogen, with the provision that the nitrogen may carry a positive charge.

Please amend claim 3 as follows:

3. (once amended) The method according to [anyone of claims 1-2]claim 1, **characterized** in that at least one of R₁₋₄ exhibit an electron withdrawing group, preferably selected among halogens such as fluorine.

Please amend claim 4 as follows:

4. (once amended) The method according to [anyone of claims 1-3]claim 1, **characterized** in that the spacer [have]has a sulphur atom next to X.

Please amend claim 5 as follows:

5. (once amended) The method according to [any one of claims 1-4]claim 1, **characterized** in that Z and Y are nitrogens, one of which binding to a hydrogen and the ligand structure being charged depending of pH.

Please amend claim 6 as follows:

6. (once amended) The method of [anyone of claims 1-5]claim 1, **characterized** in that said mixture derives from a host in which said serum albumin is human serum albumin.

Please amend claim 7 as follows:

7. (once amended) The method of [anyone of claims 1-6]claim 1, **characterized** in that said ligand is attached covalently to said matrix.

Please amend claim 8 as follows:

8. (once amended) The method of [anyone of claims 1-7]claim 1, **characterized** in that after the adsorption step said serum albumin is eluted from said affinity adsorbent and if necessary further processed.

Remarks

Claims 1-10 are pending in the instant application. Applicants have amended claims 1, 3, 4, 5, 6, 7, and 8 to more fully conform with U.S. practice and to delete multiple dependencies. A version of the claims marked up to show the amendments, as well as a clean version of the claims encompassing the amendments, is attached hereto.

Applicants respectfully assert that all amendments are fairly based on the specification, and respectfully request their entry.

Applicants believe that the claims, as amended, are in allowable form, and earnestly solicit the allowance of claims 1-10.

Respectfully submitted,



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Claims (marked-up version showing amendment(s))

[C L A I M S]

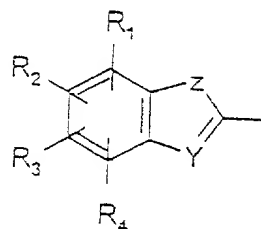
What is claimed is:

1. (once amended) [A] In a method for selectively enriching/removing a serum albumin from a mixture of other compounds by contacting said mixture with a ligand (= X), the improvement comprising said ligand
 - a) having affinity for and enabling binding of the serum albumin and
 - b) being attached via a spacer (= B) to a base matrix (= M') insoluble in the aqueous media used, the matrix with the attached ligand being represented by

M-B-X

where M is the matrix, B is the spacer and X the affinity ligand, with the provision that M may contain further groups X linked via a spacer,

characterized in that said ligand X has been selected among serum albumin-binding structures complying with the formulae



in which

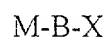
- a) the free valence bind to the spacer B;
 - b) R_{1-4} are selected from hydrogen, electron-withdrawing groups, such as halogens and lower alkyl groups (C_{1-10}) that possibly are substituted with electron withdrawing groups, such as halogens;
 - c) Z and Y are selected among oxygen, sulphur or nitrogen, with the provision that the nitrogen may carry a positive charge.
3. (once amended) The method according to [anyone of claims 1-2] claim 1, **characterized** in that at least one of R_{1-4} exhibit an electron withdrawing group, preferably selected among halogens such as fluorine.

4. (once amended) The method according to [anyone of claims 1-3]claim 1, **characterized** in that the spacer [have]has a sulphur atom next to X.
5. (once amended) The method according to [any one of claims 1-4]claim 1, **characterized** in that Z and Y are nitrogens, one of which binding to a hydrogen and the ligand structure being charged depending of pH.
6. (once amended) The method of [anyone of claims 1-5]claim 1, **characterized** in that said mixture derives from a host in which said serum albumin is human serum albumin.
7. (once amended) The method of [anyone of claims 1-6]claim 1, **characterized** in that said ligand is attached covalently to said matrix.
8. (once amended) The method of [anyone of claims 1-7]claim 1, **characterized** in that after the adsorption step said serum albumin is eluted from said affinity adsorbent and if necessary further processed.

Claims (clean version encompassing amendments)

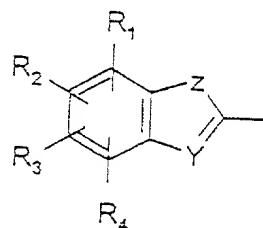
What is claimed is:

1. (once amended) In a method for selectively enriching/removing a serum albumin from a mixture of other compounds by contacting said mixture with a ligand (= X), the improvement comprising said ligand
 - a) having affinity for and enabling binding of the serum albumin and
 - b) being attached via a spacer (= B) to a base matrix (= M') insoluble in the aqueous media used, the matrix with the attached ligand being represented by



where M is the matrix, B is the spacer and X the affinity ligand, with the provision that M may contain further groups X linked via a spacer,

characterized in that said ligand X has been selected among serum albumin-binding structures complying with the formulae



in which

- a) the free valence bind to the spacer B;
 - b) R₁₋₄ are selected from hydrogen, electron-withdrawing groups, such as halogens and lower alkyl groups (C₁₋₁₀) that possibly are substituted with electron withdrawing groups, such as halogens;
 - c) Z and Y are selected among oxygen, sulphur or nitrogen, with the provision that the nitrogen may carry a positive charge.
2. The method according to claim 1, **characterized** in that contact between the mixture and the media M-B-X is done in an aqueous media having a pH at which the -B-X carries a positive charge.

3. (once amended) The method according to claim 1, **characterized** in that at least one of R1-4 exhibit an electron withdrawing group, preferably selected among halogens such as fluorine.
4. (once amended) The method according to claim 1, **characterized** in that the spacer has a sulphur atom next to X.
5. (once amended) The method according to claim 1, **characterized** in that Z and Y are nitrogens, one of which binding to a hydrogen and the ligand structure being charged depending of pH.
6. (once amended) The method of claim 1, **characterized** in that said mixture derives from a host in which said serum albumin is human serum albumin.
7. (once amended) The method of claim 1, **characterized** in that said ligand is attached covalently to said matrix.
8. (once amended) The method of claim 1, **characterized** in that after the adsorption step said serum albumin is eluted from said affinity adsorbent and if necessary further processed.
9. A method for screening ligands structures that, when attached to an affinity matrix, selectively bind albumin, characterized that water-soluble compounds that exhibit a benzene ring fused to a 5-membered heterocycle containing two or three heteroatoms, preferably two, selected from nitrogen, oxygen and sulphur after having been attached to a matrix, preferably in the 2-position, are screened for selective binding to albumin.

10. The method of claim 9, characterized in that screening is taking place in aqueous solutions at a pH at which the ligand structure including and spacer binding to the matrix is positively charged.

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REMOVAL/PURIFICATION OF SERUM ALBUMINS

Technical field

The present invention concerns a method for the selective separation/removal of a mammalian serum albumin from a solution containing a mixture of proteins in order to obtain a solution/preparation that is substantially devoid of one or more of the other proteins, for instance immunoglobulins and/or serum albumin of species origin other than the separated/removed serum albumin.

Technical background

For a long time there has been a large demand for purified mammalian serum albumins, for instance serum albumin of human or bovine origin (HSA and BSA, respectively). For HSA this has mainly depended on its therapeutic use as a plasma volume expander. Originally serum albumins were obtained from sera/plasma of the appropriate species origin. For some years the focus has been to produce serum albumins recombinantly, in particular HSA. For bacterially produced recombinant forms, it has become urgent to remove host cell contaminants because they may be hazardous in vivo to mammals. It has become apparent that preparing HSA in transgenic animals should be beneficial, for instance in transgenic cows. This latter alternative, however, has the drawback that HSA will be present in mixture with the HSA analogue of the host animal (for instance with BSA if HSA is produced in cows). This has created novel purification problems, for instance the specific removal of BSA from HSA.

Various forms of affinity binding including ion exchange binding have earlier been applied to the purification of serum albumins. For affinity binding the general goal has been to find chromatographic media (ligand attached to a chromatographic base matrix) that provide sufficient specificity in order to remove

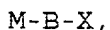
either predetermined contaminants or the serum albumin desired as the end product from complex mixtures. The term "binding" includes chromatographic as well as batch-wise alternatives.

Illustrative examples of ligands previously used and having selectivity for serum albumins are given by Theodore Peters in All about Albumin - Biochemistry, Genetics and Medical Applications, (Ed. Theodore Peters, Jr., Academic Press (1996) pages 77-126. Among low molecular weight ligands there may be mentioned certain dyes of rather complex structure, for instance Cibacron Blue.

WO 9602573 discloses a method for selectively purifying human serum albumin from whole milk or whey by a process comprising an affinity chromatographic step utilizing triazine dye molecules as affinity ligands.

WO 9808603 discloses a number of aromatic compounds that can be used as general protein binders under the appropriate conditions. The compounds includes 2-mercapto-benzothiazoles among several others. Serum albumin is used as one of many model compounds.

The methods concerned in the present invention generally provide that a mixture of compounds, in particular proteins, containing a serum albumin is contacted with a ligand that has affinity for the serum albumin and that is attached to a base matrix. The conditions are adjusted such that the serum albumin becomes selectively bound (adsorbed) to the ligand. The matrix-bound affinity ligand may be represented by the formula I



where M is the matrix, X the affinity ligand (ligand structure) and B a spacer group through which X is attached to the matrix. M may contain several groups X linked through spacers that may be equal or unequal to B.

The expression "a ligand that is attached to a matrix" includes also ligands that are possible to attach to a matrix after having

bound a serum albumin.

Objectives of the invention

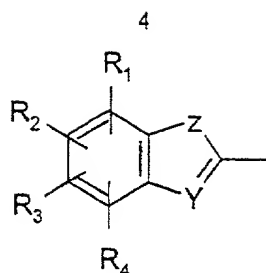
The first objective of the invention is to provide novel
5 affinity methods for the selective removal of a serum albumin
from a mixture of proteins in order to produce the serum albumin
in substantially pure form or a preparation essentially free of
the removed serum albumin.

A second objective is to provide an affinity method as defined
10 above which is selective for a serum albumin that exist in
mixture with one or more other serum albumins.

The invention

It has now been discovered that there are compounds that
15 selectively bind albumin to be found among bicyclic compounds in
which a five-membered heterocycle is fused with a benzene ring.
The heterocycles concerned contain two or three heteroatoms
selected from oxygen, nitrogen and sulphur. It has also been
found that this type of albumin-binding compounds can be used as
20 affinity ligands attached to matrices for the
purification/removal of serum albumins from liquids. It has also
been found that this ability to bind albumin may differ depending
on the species origin of the serum albumin.

The inventive method thus has the characterizing feature that
25 the ligand X defined above has been selected from the group of
compounds defined in the preceding paragraph and linked to an
affinity matrix M through a spacer B. In particular X complies
with the structure according to formula II:



The free valence binds to the spacer B.

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R_1 , R_2 , R_3 and R_4 are selected from hydrogen, electron-withdrawing groups and lower alkyl groups (C_{1-10}) that possibly are substituted with electron withdrawing groups and/or hydrophilic groups. Particular important alkyl groups (R_{1-4}) contain 1-3, preferably one, carbon atom. Particular important electron withdrawing groups are halogens, such as F, Cl and Br, which also may enhance a hydrophilic character of the ligand. A well known hydrophilic group is hydroxy ($HO-$) that, like the halogens, when present on an alkyl group will enhance the hydrophilic character of the latter. Particular interesting groups R_{1-4} comprise C_{1-10} , such as C_{1-4} , alkyl groups substituted with 1-3 halogen atoms, preferably fluorine, on carbons at none, one or two atoms distance from the ring structure. One, two or three of R_{1-4} may be different from hydrogen, although it is believed that in the preferred ligands X one or two of R_{1-4} should not be hydrogen. Non-hydrogen groups R_{1-4} may be located in the 4-, 5-, 6- and/or 7-position with preference for positions 4 and/or 6.

Z and Y are heteroatoms selected from nitrogen, oxygen and sulphur, possibly carrying one or two organic groups or hydrogens in addition to the heterocyclic ring they are part of. Normal valence rules for stable compounds apply, i.e. the nitrogen may carry one or two, the sulphur one and the oxygen none additional group(s). The organic groups may be selected according to the same principles as R_{1-4} , with preference for smaller alkyl groups and hydrogen. In the heterocyclic ring a nitrogen atom and a

sulphur atom can be positively charged if they bind to other atoms or groups via four and three valencies, respectively.

In the preferred ligand structures X, Z and Y are NR'R'' and NR''', respectively. R', R'' and R''' are selected according to the same principle as R₁₋₄ or is a free electron pair, with the proviso that a selected R₁₋₄ group should provide a saturated sp³-hybridised carbon atom binding directly to the heterocyclic ring. The preferred ligand structures of this embodiment comprise that at most one of R' and R'' is a free electron pair or a hydrogen. In this latter embodiment R''' is typically a free electron pair.

The general idea is that the useful types of ligands shall have a pronounced hydrophilicity in order to increase the desired selectivity for serum albumins and to minimize binding of other kinds of proteins via hydrophobic interactions. It is therefore believed that ligand structures that selectively bind albumin will be found by selecting or screening among water-soluble organic compounds that

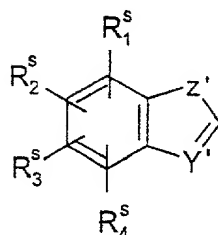
- a) comprise a benzene ring fused to a five-membered heterocycle containing two or three, preferably two, heteroatoms selected from nitrogen, oxygen and sulphur and
- b) have been attached to a matrix M via a spacer B replacing a group or atom at any of the positions 1, 2, 3, 4, 5, 6 and 7

for selective binding to serum albumin. Once a compound according to formula II has been pretested or prescreened and found capable of binding selectively a serum albumin, this finding may be used by anyone to selectively remove the serum albumin as discussed above.

Particularly interesting compounds have two ring heteroatoms which are present in position 1 and 3, respectively, such as those compounds complying with the general structure given in formula III. Attachment to a matrix is preferably at the 2-

6

position of the ring system. The screening methodology may be of the same kind as given in the Experimental Part.



Z' and Y' are heteroatoms selected according to the same rules as for Z and Y. In case they comprise nitrogen as the heteroatom they may be NR_s'R_s' and NR_s'', respectively. The substituents R_s¹⁻⁴ and R_s', R_s' and R_s' and the groups/atoms Y' and Z' are selected such that the compound III is soluble in water at a pH-value in the interval 1-11, preferably 1-7 (25°C). By solubility in water is meant at least 0.01 mg/ml, with preference for at least 0.1 mg/ml. Typically this means that, when one or more of R_s¹⁻⁴ (for instance as defined for R₁₋₄ above) are alkyl groups with a relatively long carbon chain, there should also be present one or more hydrophilic substituents, for instance hydroxy, amino, carboxy and other related substituents (ether, thioether, amido, ester etc), on the chain. The chain may also be broken by one or more heteroatoms, such as sulphur (thioether), oxygen (ether) or nitrogen atoms (secondary, tertiary or quaternary amino). Preferably the compound (and the ligand structure) should exhibit a charge (positive or negative) in a part of this pH interval (positive in the lower part and/or negative in the upper part or positive all throughout the pH-interval 1-10. It is believed that screening for binding preferentially should take place at a pH

where the ligand structure have a charge (≥ 0 , preferably > 0). This includes that the charge may be located at a nitrogen in any of the groups R_{1-4}^s and R_s' , R_s'' and R_s''' or even in the spacer B.

- 5 At the priority the preferences with respect to R_{1-4}^s and R_s' , R_s'' and R_s''' were the same as for R_{1-4}^s , R' , R'' and R''' and Y and Z.

The hydrogen (H) in position 2 may be replaced by the part of the spacer B that is next to X in formula I, for instance a part
10 containing a spacer chain of at most 10 atoms in length.

The matrix and the attachment of the ligand to the matrix.

In the preferred modes of the invention the ligand is attached to a base matrix that is insoluble in the aqueous media used.

- 15 Such matrices often are based on polymers that expose a hydrophilic surface to the aqueous media used, i.e. expose hydroxy (-OH), carboxy (-COOH), carboxamido (-CONH₂, possibly in N- substituted forms), amino (-NH₂), oligo- or polyethylenoxy groups on their external and, if present, also on internal
20 surfaces. Typically the matrices are of the same kind as those normally used as chromatographic matrices. The polymers may, for instance, be based on polysaccharides, such as dextran, starch, cellulose, pullulan, agarose etc, which if necessary have been crosslinked, for instance with bisepoxides, epihalohydrins,
25 1,2,3-trihalo substituted lower hydrocarbons, to provide a suitable porosity and rigidity. The matrices may also be based on synthetic polymers, such as polyvinyl alcohol, poly hydroxyalkyl acrylates, poly hydroxyalkyl methacrylates, poly acrylamides, polymethacrylamides etc. In case of hydrophobic polymers, such as
30 those based on divinyl and monovinyl substituted benzenes, the surfaces of the matrices are often hydrophilized to expose hydrophilic groups as defined above.

The matrices may also be of inorganic nature, e.g. silica, zirkonium oxide etc.

Physically the insoluble matrices may be in the form of porous monoliths or in beaded/particle form that can be either porous or non-porous. Matrices in beaded/particle form can be used either as a packed bed or in suspended form. Suspended forms include so called classified expanded beds and pure suspensions in which the particles/beads are moving round completely. In case of monoliths, packed bed and classified expanded beds, the separation procedure may be classified as a normal chromatography with a concentration gradient of adsorbed molecules being established along the flow direction. In case of pure suspension the separation procedure will be in the batch wise mode.

For suspensions, the beads/particles may contain a densifying filler material that will permit higher flow rates in case of classified expanded beds and facilitate sedimentation of the beads/particles after adsorption. See for instance WO-A-9717132 (Amersham Pharmacia Biotech AB) and WO-A-9200799 (Upfront Chromatography).

The matrices may also comprise so called extenders carrying several groups -B-X. Extenders are often polymeric providing attachments of several ligands per spacer. Extenders are often hydrophilic and called tentacles etc. See for instance International Patent Application PCT/SE98/00189 (Amersham Pharmacia Biotech AB).

A spacer is typically introduced to improve the availability of the ligand and/or facilitate the chemical coupling of the ligand to the matrix. Spacers often comprise a hydrocarbon chain that has a length between 1-50 atoms. The hydrocarbon chain may be straight, branched or cyclic and optionally broken with one or more ether oxygen or amino nitrogen atoms and/or optionally substituted with one or more hydroxy, lower alkoxy, or amino

group ($-\text{NH}_2/\text{NH}_3^+$, where each hydrogen may be replaced with a lower alkyl or a lower acyl group). By lower alkyl or lower acyl group is primarily intended C_{1-10} alkyls/acyls. The spacer group may also, depending to coupling methodology, comprise ester, amido, thioether, etc groups that have the sufficient hydrolytic stability. This latter groups may be present in the spacer either alone or combined with each other and/or with the appropriate hydrocarbon chain(s).

In the spacer the atom binding directly to the ligand structure X should preferably be a sulphur atom, preferably bound to a saturated carbon atom in the spacer. In other alternatives of the spacer, the atom next to the ligand structure X may be an oxygen, a nitrogen (amino or amido nitrogen), a carbon (for instance a saturated carbon, carbonyl carbon etc). Typical saturated carbon atoms are only binding carbons and/or hydrogen.

The spacer, in particular the part closest to ligand structure X, may influence the ability to bind to serum albumin. The discussion above therefore primarily indicates the group of spacers that is to be screened for finding the optimal spacer for various ligand structures.

It can be envisaged that the ligand potentially also will be attached to the matrix by methods involving non-covalent bonding, such as physical adsorption or biospecific adsorption. In these cases the ligand may be covalently attached, via a spacer structure, to some type of carrier molecule that provides the ligand with sufficient adsorption ability relative adsorbing structures in the matrix. The carrier molecule and the adsorbing structures are in these cases regarded as being part of the matrix (M'). Pairs of carrier molecules and adsorbing structures may be represented by the biotin-streptavidin system.

As a potential alternative, the ligand may be in soluble form that subsequent to binding to a serum albumin is insolubilized.

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This may be accomplished, for instance, by having the ligand conjugated to biotin and insolubilizing by contacting the formed complex between the serum albumin and the soluble ligand-biotin conjugate with a streptavidin-matrix. The spacer in this mode will
5 be the grouping linking the ligand to biotin.

The sample containing the serum albumin to be removed/purified

The serum albumin to be removed and/or purified typically exists in mixture with other proteins and/or biomolecules.

10 Illustrative examples are blood preparations (such as plasma and serum), fermentation liquids obtained from cultured host cells that have been transformed to express a serum albumin, biological fluids obtained from transgenic mammals transformed to produce a serum albumin of another species, and also various working up
15 preparations derived from any one of these types of liquids. Important biological fluids from transgenic mammals are milk and milk fractions such as whey. In case of liquids derived from transgenic animals the liquids will often contain also the normal serum albumin of the species concerned.

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Procedural steps for binding

During the adsorption step the conditions are selected so as to promote binding between the ligand and the serum albumin intended. At the same time the conditions are selected such that
25 binding between the ligand and other proteins present is suppressed, for instance hydrophobic interactions.

For pH this means that the pH-value shall be selected above the value at which serum albumin unfold. This value in turn may depend on ionic strength and temperature. The uppermost end of
30 the pH is limited at the pH value at which hydrolytic degradation becomes significant. As a general rule of thumb, the appropriate pH values will therefore be found in the range 4-10. However, our

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present findings suggest that the binding may require a positive charge on the ligand structure. The very broad range 4-10 may therefore only be applicable to ligand structures, including the spacer B, carrying a pH-independent charge, for instance Z being a quaternary ammonium group. In case one or more of Z, Y, R^s₁₋₄ and R_s['], R_s^{''} and R_s^{'''} or B comprise a nitrogen that can be protonated and deprotonated in water, the ligand structure X plus the spacer B may exhibit a positive charge which is pH-dependent. The lower part of the pH interval 4-10 will then be applicable, for instance pH 4-7 or even lower such as 4-6.

The ionic strength should be selected in the interval corresponding to 0-3 M NaCl. Ionic strengths at the upper part of this interval are likely to promote binding via hydrophobic interactions. This means that albumin binding will be promoted at higher ionic strengths, but simultaneously also proteins with pronounced hydrophobic regions will bind more efficiently thereby decreasing the selectivity for serum albumins. The optimal ionic strength will therefore, among others, depend on what is to be removed from what. The temperature is typically selected in the interval 0-40°C with preference for 4-37°C.

The optimal values of pH, temperature and ionic strength will depend on the species origin of the serum albumin to be removed/purified, contaminants, the ligand structure attached to the matrix etc.

After adsorption, the serum albumin may be further worked up, for instance by first desorbing the serum albumin and subsequently subjecting it to further adsorption steps, for instance on a cation or anion exchanger, an hydrophilic matrix exhibiting hydrophobic groups (HIC-media) etc. Suitably desorption conditions may include change of pH, of ionic strength, of temperature and/or addition of compounds interfering with binding. Compounds that interfere with the binding may

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mimick the ligand structure X present on the matrix. It may, for instance, be a compound according to formula III possibly also exhibiting a relevant part of the spacer B. The main rules should then of course be not to change the conditions so that the serum albumin becomes irreversibly denatured. The adsorption step may be preceded by other steps, which for instance may be based on ion exchange or immune ligands. Illustrative examples of steps that can be combined with the present inventive adsorption step are given in US 5440018; US 5521287 and EP 699687 (all in the name of Green Cross Corp).

The goal of the full separation processes is to obtain serum albumin preparations intended for use in vivo. This means that the preparation should contain < 0,01 %, such as < 0,001 % proteins that are heterologous to the species to which the preparation is to be administered. HSA preparations, for instance, that are to be used in humans shall contain < 0,01 %, such as < 0,001 % BSA. All percentages are w/w. This means that in order to reach a therapeutically acceptable purity the serum albumin preparations obtained directly from the adsorption step of the present invention often will need to be combined with further working up steps, either prior or subsequent to the inventive adsorption step.

The invention is further defined in the appended claims and will now be further described by experimental support regarding the most preferred fragment.

EXPERIMENTAL PART

The purpose of the study was to immobilize a number of selected heterocycles condensed to a benzene ring (benzo compounds) to an epoxy activated matrix (Sepharose 4FF, Amersham Pharmacia Biotech

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AB, Uppsala, Sweden) and test them for binding to human serum albumins and IgG in PBS buffer at pH 7. In order to facilitate attachment to the matrix the compounds were selected to carry a thiol group (at the 2-position). The ligand structures were:

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No	Ligand structure X	Manufacturer of coupled compound	Subst. (mole/ml gel)	Vt ml
1	Benzimidazol-2-yl	Aldrich	15.1	1.2
2	5-methyl-benzimidazol-2-yl	Aldrich	14.4	1.2
3	Benzothiazol-2-yl	Aldrich	8.4	
4	Benzoxazol-2-yl	Aldrich	12.2	1.2
5	4-bromo-6-(trifluoromethyl)-benzimidazol-2-yl	Maybridge	15.1	1.2
6	6-ethoxy-benzothiazol-2-yl	Aldrich	9.6	1.1
7	5-nitro-benzimidazol-2-yl	Aldrich	15.1	1.2
8	6-nitro-benzothiazol-2-yl	TCI-GB	6.8	1.2
9	5-chloro-benzothiazol-2-yl	Aldrich	9.6	1.2
10	5-methoxy-benzimidazol-2-yl	Aldrich	14.7	1.1
11	4-chloro-6-(trifluoromethyl)-benzimidazol-2-yl	Maybridge	14.3	1.2
12	5-chloro-1-	Maybridge	18.2	1.2

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	isopropyl- benzimidazol-2-yl			
13	5,6-dichloro- benzimidazol-2-yl	Maybridge	15.1	1.2
14	5-chloro-6-fluoro- benzimidazol-2-yl	Maybridge	4.9	1.1

Test proteins: Human Serum Albumin (HSA) (Sigma), Bovine Serum Albumin (BSA) (Sigma) and IgG human normal (Pharmacia AB = Pharmacia & Upjohn, Stockholm, Sweden).

Buffers used in binding experiments:

1. PBS pH 7
2. 20 mM citrate buffer. Dissolve 2.52 g citric acid monohydrate and 1.90 g citric acid monosodium salt in 1000 ml MilliQ water.
3. 2 M NaCl. Dissolve 116.88 g in 1000 ml MilliQ.
4. West. Add 6.8 g sodium acetate to 840 ml MilliQ water. Adjust pH with hydrochloric acid. Add 47.9 lactose, 0.44 g calcium chloride and 20 ml 2M NaCl. Add MilliQ water to 1000 ml.

Test solutions:

HSA (2 mg/ml in PBS)

HSA (2 mg/ml in West)

BSA (2 mg/ml in West)

IgG (1.5 mg/ml in PBS; dilute 1 ampoule (165 mg/ml) to 100 ml)

Synthesis of affinity media: 100 ml of drained gel (Sephacrose FF (Amersham Pharmacia Biotech AB, Uppsala) were washed on a glass filter with 400 ml of distilled water and transferred to a round bottomed glass flask together with 5.5 g of sodium hydroxide dissolved in 30 ml of distilled water. The mixture was kept at

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35°C. 80 ml of 1,4-bis(epoxypropoxy)-butan were added under vigourous stirring. After modest stirring for two hours the gel was washed with 1.5 l of distilled water.

In each synthesis 1 mmole of the compound to be coupled
5 (compounds 1-15) was dissolved in 5 ml DMSO. The epoxy gel was washed with DMSO followed by addition of the ligand solution to 5.0 g of gel. 2 drops of 45% sodium hydroxide solution were added to each reaction flask (pH about 11). The reaction vessels were incubated at 40°C for 60 hours. The gels were filtered on glass
10 funnels and were washed with the following solvents: 1. DMSO; 2. DMSO/water (1:1); 3. water; 4. THF; 5. water; 6. ethanol; and 7. water. The substitution degree was determined by elemental analysis.

15 Packing of columns: The gel slurry was poured into the column and packed by sucking the solution out with a syringe attached to the column outlet. The adapter was mounted on the column and the gel was equilibrated with about 10 columns of PBS buffer.

20 Binding of HSA or human IgG in PBS pH 7 - elution with citrate pH 3: The column was equilibrated with PBS pH 7 after which 2.000 ml HSA (2 mg/ml) or IgG (16.5 mg/ml) was applied. The column was eluted first with buffer of pH 7 and then with buffer of pH 3 (citrate; wash buffer).

25 Binding of HSA and BSA in West pH 4.6 - step wise elution with PBS pH 7 and citrate pH 3 : The column was equilibrated in West buffer pH 4.6 and 2.000 ml HSA (2 mg/ml) or BSA (2 mg/ml) was applied. Elution was made West pH 4.6 then with PBS pH 7 and
30 finally with citrate pH 3.0 (wash).

Results:

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Binding of HSA or human IgG in PBS pH 7 - elution with citrate pH 3: Based on conventional ways of interpreting the chromatogram recorded, none of the ligand structures showed binding to IgG or HSA. The buffer was the standard for this type of experiments with serum albumin (PBS pH 7). In spite of these negative results the present inventors went further on and analysed in more detail the shape and position of the peaks of the chromatograms. It was found that for HSA the chromatograms sometimes looked somewhat different for different ligand structures. All chromatograms for IgG looked the same and the position of the eluted IgG suggested no interaction/binding. For seven (3, 4, 6, 8, 9, 12 and 13) out of the 14 tested ligand structures the chromatograms looked the same with the HSA peak located at the same elution volume and having the same shape.

The HSA chromatograms from the other seven columns (1, 2, 5, 7, 10, 11 and 14) could be paired, two by two, due to a similarity and/or retardation of the peaks compared to the peaks for the previously discussed ligand structures. The ligand structures 1 and 2 showed two peaks in the flow through and ligand structures 7 and 14 showed peaks with a shoulder. At least the latter result together with the retardation suggest an interaction with the media. Ligand structures 5 and 11 gave retarded peaks that were tailed. A very small elution peak and a small peak when reequilibrating the column could also be seen. The tailing and retardation suggested some type of weak interaction with the ligand structure on the media. Ligand structure 10 gave a small peak when reequilibrating the column.

Binding of HSA or BSA in West pH 4.6 - step wise elution with PBS pH 7 and citrate pH 3: The column with ligand structure 11 was studied with HSA and with BSA. The chromatograms indicated that all HSA applied and a part of the BSA applied were bound in West pH 4.6 (no HSA was eluted until PBS pH 7 was applied, one

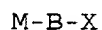
17

portion/peak of BSA eluted with West 4.6 and another with PBS pH 7).

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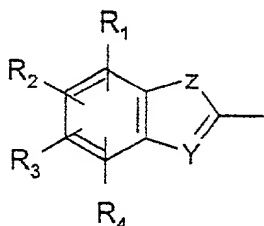
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C L A I M S

- 1 A method for selectively enriching/removing a serum albumin
from a mixture of other compounds by contacting said mixture
5 with a ligand (= X), said ligand
- a) having affinity for and enabling binding of the serum
albumin and
 - b) being attached via a spacer (= B) to a base matrix (= M')
- 10 insoluble in the aqueous media used, the matrix with the
attached ligand being represented by



where M is the matrix, B the spacer and X the affinity
ligand, with the provision that M may contain further groups
X linked via a spacer,

15 characterized in that said ligand X has been selected among
serum albumin-binding structures complying with the formulae



- 20 in which
- a) the free valence bind to the spacer B;
 - b) R_{1-4} are selected from hydrogen, electron-withdrawing groups,
such as halogens and lower alkyl groups (C_{1-10}) that possibly
are substituted with electron withdrawing groups, such as
25 halogens;
 - c) Z and Y are selected among oxygen, sulphur or nitrogen, with
the provision that the nitrogen may carry a positive charge.

2. The method according to claim 1, characterized in that contact between the mixture and the media M-B-X is done in an aqueous media having a pH at which the -B-X carries a positive charge.

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3. The method according to anyone of claims 1-2, characterized in that at least one of R1-4 exhibit an electron withdrawing group, preferably selected among halogens such as fluorine.

- 10 4. The method according to anyone of claims 1-3, characterized in that the spacer have a sulphur atom next to X.

- 15 5. The method according to any one of claims 1-4, characterized in that Z and Y are nitrogens, one of which binding to a hydrogen and the ligand structure being charged depending of pH.

- 20 6. The method of anyone of claims 1-5, characterized in that said mixture derives from a host in which said serum albumin is human serum albumin.

7. The method of anyone of claims 1-6, characterized in that said ligand is attached covalently to said matrix.

- 25 8. The method of anyone of claims 1-7, characterized in that after the adsorption step said serum albumin is eluted from said affinity adsorbent and if necessary further processed.

- 30 9. A method for screening for ligands structures that, when attached to an affinity matrix, selectively bind albumin, characterized that water-soluble compounds that exhibit a benzene ring fused to a 5-membered heterocycle containing two

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or three heteroatoms, preferably two, selected from nitrogen, oxygen and sulphur after having been attached to a matrix, preferably in the 2-position, are screened for selective binding to albumin.

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10. The method of claim 9, characterized in that screening is taking place in aqueous solutions at a pH at which the ligand structure including and spacer binding to the matrix is positively charged.

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DESIGN
PATENT APPLICATION
(37 CFR 1.63)**

☐ Declaration Submitted with Initial Filing
OR
☒ Declaration Submitted after Initial Filing (surcharge (37 CFR 1.16 (e)) required)

Attorney Docket Number PU-9843

First Named Inventor Tor Regberg

COMPLETE IF KNOWN

Application Number 09 /869,023

Filing Date 20-Jun-2001

Group Art Unit To be assigned

Examiner Name To be assigned

As a below named inventor, I hereby declare that:

My residence, post office address, and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

Removal/Purification of Serum Albumins

the specification of which (Title of the invention)

☐ is attached hereto
OR

☒ was filed on (MM/DD/YYYY) 06/20/2001 as United States Application Number or PCT International

Application Number 09/869,023 and was amended on (MM/DD/YYYY) (if applicable)

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment specifically referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in 37 CFR 1.56.

I hereby claim foreign priority benefits under 35 U.S.C. 119(a)-(d) or 365(b) of any foreign application(s) for patent or inventor's certificate, or 365(a) of any PCT international application which designated at least one country other than the United States of America, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or of any PCT international application having a filing date before that of the application on which priority is claimed.

Prior Foreign Application Number(s)	Country	Foreign Filing Date (MM/DD/YYYY)	Priority Not Claimed	Certified Copy Attached?	
				YES	NO
9804465-4	Sweden	12/22/1998	<input type="checkbox"/>	<input type="checkbox"/>	<input checked="" type="checkbox"/>
			<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
			<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
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U.S. Parent Application or PCT Parent Number	Parent Filing Date (MM/DD/YYYY)	Parent Patent Number (if applicable)
PCT/EP99/10123	12/20/1999	

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Name of Sole or First Inventor:

☐ A petition has been filed for this unsigned inventor

Given Name (first and middle [if any])		Family Name or Surname	
Tor		Regberg	
Inventor's Signature	Date		
Residence: City	State	Country	Citizenship
Post Office Address			
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ADDITIONAL INVENTOR(S) Supplemental Sheet

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Given Name (first and middle (if any))		Family Name or Surname	
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			07/11/01
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Given Name (first and middle (if any))		Family Name or Surname	
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